Spawner-isolated mortality virus - Real Time DNA

50 / 100 /150 tests (Ready to use kit)

OneVetqPCR-realtime™ SMV DNA Real Time PCR Kit is a screening assay for a rapid and accurate detection of Spawner-isolated mortality virus.

Principles of the test:

One Step PCR Kits provide components for "onestep" real time PCR detection in a convenient format that is compatible with both rapid and standard qPCR cycling conditions.
The One Universal qPCR DNA Master Mix include all reagents for an optimized qPCR.
The Spawner-isolated mortality virus specific primer and probe mix are provided in the kit and these can be detected through your real time Platform by the 5' nuclease PCR detection method. During PCR amplification, forward and reverse primers hybridize to the Spawner-isolated mortality virus target genomic DNA generated. Fluorogenic probe is included in the same reaction mixture which consists of a DNA probe labeled with a 5-reporter FAM and 3-quencher which can be detected through green channel.
To confirm extraction of a valid biological template an Internal control primer and probe mix is included, consists of a DNA probe labeled with a 5-reporter HEX and a 3-quencher which hybridize inside a specific housekeeping endogenous target gene. During PCR amplification, the probe is cleaved and the reporter dye and quencher are separated. As a result, a fluorescence increase can be detected on a range of real time PCR platforms through yellow channel.
Our kits also include Positive and Negative Control which are details in FAQ section.

Principle and use:

This amplification kit has been manufactured to detect Spawner-isolated mortality virus in real time PCR.
This is a possibility absolute quantification or qualitative assay.
Real time PCR is based on fluorogenic dyes. Ct value between 12 - 36 should be taken positive. Value between 36 - 40 Ct should be taken as marginal positive. Ct above 40 must be considered as negative **(for more details see Table 5).**
This kit needs DNA as a template which can be isolated from blood, serum, faeces, respiratory fluid, cerebrospinal fluid, digestive system, tissue, Heptopancreas, Gills, Pleopods, Cloacal, Egg Yolk, Milk, swabs, Lee, bacterial culture, cell lines and others. We discarded use of affinity columns because a lot report that indicate purification problems due to the lipids present in the biological samples quickly clog the column decreasing its performance.

Table 1. Kit Components:

Reactions Tubes	50 test	100 test	150 test
Universal qPCR Master Mix	(1 vial)	(1 vial)	(1 vial)
Primer, Probes and Internal Control Universal Mix	(1 vial)	(1 vial)	(1 vial)
SMV Positive Control	(1 vial)	(1 vial)	(1 vial)
SMV Negative Control	(1 vial)	(1 vial)	(1 vial)
PCR grade Water	(1 vial)	(1 vial)	(1 vial)

Table 2. Instrument Compatibility:

* ABI 7300	* LightCycler 2.0
* ABI 7500FAST	* LightCycler 480
* ABI 7900	* Mastercycler® ep realplex
* AB Step One	* Mx3000P QPCR System
* AB Step One Plus	* Mx3005P QPCR System
* Agilent Mx3005P	* RotorGene 3000
* CFX96 & CFX384	* RotorGene 6000
* ExiCyclerTM 96	* RorotGeneQ
* iQ5 & MyiQ Cycler	* SLAN® Real-Time PCR
* Illumina Eco	* Smartcycles II
* LightCycler Nano	* Applied 7300 and 7500

Procedure:

Please read through the entire procedure before starting.

Before Starting

- Pulse-spin each tube in a centrifuge before opening.
- Homogenize the solutions for 5 seconds prior to pipetting
- You must consider use different tips in order to avoid cross contamination.
- Use only sterile, RNAses, DNAases and pyrogens free tips.

Step 1

Prepare a Master mix according to the reaction table.

Table 3. Reaction components for PCR

Reaction Tubes	Sample and Internal Control	Positive Control	Negative Control
Universal qPCR Master Mix	7,5 µL	7,5 µL	7,5 µL
Primer, Probes and Internal Control Universal Mix	0,3 µL	0,3 µL	0,3 µL
PCR grade Water	3,7 µL	3,7 µL	3,7 µL
DNA Sample	3,5 µL		
SMV Positive Control		3,5 µL	
SMV Negative Control			3,5 µL
Total Volume	15 µL	15 µL	15 µL

Step 2

Place the tubes in a thermal cycler and perform One Step qPCR according to the program outlined in Table 2.

Table 4. Recommended PCR Cycling table

Cycle	Steps	Time	Temp.(°C)
1 Cycle	Activation Step	1 min	95°C
40 Cycles	Denaturation	30 seg	95°C
	Annealing	30 seg	58°C
	Extension	30 seg	72°C
	Final Step	∞	4°C

Interpretation of the test

1) Qualitative analysis:

Ct (Threshold cycle) value of each sample can be read as follows.

Table 5. Ct value result

Ct value	Result
0 - 11	**Negative**
12 - 36	Positive
36 - 40	Marginal Positive
> 40	**Negative**

* Is important mentioned that Ct value over 40 is considered Negative result. If Ct value is in a 12 - 36 range, it must be considered as Positive result. This is depending of the sample initial concentration used for each reaction. You should consider that sample real concentration could be modify by the sample purity when this is quantifier
* For more technical information you must request the quality control for each kits. Also you can request more information writing to our email info@gentaur.com

2) Quantitative analysis:

Table 6. Preparation of standard curve dilution series. SMV positive control:

Average Positive Control Concentration	
SMV	See Quality Control

Standard curve	Preparation series a fresh dilution	Concentration	Copy Number
Tube N°1	2uL SMV Positive Control (0,1 ng/µL) + 18 µL of PCR grade Water	See quality control	See quality control
Tube N°2	2uL Tube N°1 + 18 µL of PCR grade Water	See quality control	See quality control
Tube N°3	2uL Tube N°2 + 18 µL of PCR grade Water	See quality control	See quality control
Tube N°4	2uL Tube N°3 + 18 µL of PCR grade Water	See quality control	See quality control
Tube N°5	2uL Tube N°4 + 18 µL of PCR grade Water	See quality control	See quality control
Tube N°6	2uL Tube N°5 + 18 µL of PCR grade Water	See quality control	See quality control
Tube N°7	2uL Tube N°6 + 18 µL of PCR grade Water	See quality control	See quality control

Important Note: Don't forget to Homogenize the tubes.
* We will send a Quality Control report for each purchase.
** For reaction mix you must use Universal qPCR Master Mix.
*** If you want to obtain less DNA copies you must include a new dilution tube (Tube N° 8). Note: Final DNA copy number will depend of the DNA concentration (you can see it in Quality Control Report).

Table 7. Standard curve set up

	Tube A	Tube B	Tube C	Tube D	Tube E	Tube F	Tube G
Universal qPCR Master Mix	7,5 µL	7,5 µL	7,5 µL	7,5 µL	7,5 µL	7,5 µL	7,5 µL
Primer, Probes and Internal Control Mix	0,3 µL	0,3 µL	0,3 µL	0,3 µL	0,3 µL	0,3 µL	0,3 µL
PCR grade Water	3,7 µL	3,7 µL	3,7 µL	3,7 µL	3,7 µL	,7 µL	3,7 µL
Tube N° 1 (Positive Control)	3,5 µL						
Tube N° 2		3,5 µL					
Tube N° 3			3,5 µL				
Tube N° 4				3,5 µL			
Tube N° 5					3,5 µL		
Tube N° 6						3,5 µL	
Tube N° 7							3,5 µL
Total Volume	15 µL	15 µL	15 µL	15 µL	15 µL	15 µL	15 µL

2.1.-Assess the Ct value when amplification curve of Standard tube 1, 2, 3, 4, 5, 6 passes the threshold line. However, four tubes are sufficient for standard curve. (tube1-tube4).
2.2.- Calculate quantitative value to compare with Ct value of unknown samples and curve of Standard tube 1, 2, 3, 4, 5, 6.
2.3.- When you visualized result in the Real Time PCR platform you must see just one amplification curve for Positive Control. You must not see an Internal Control amplification curve.

3) Test validation:

3.1.- Each Ct value standard should be as follows.
Standard 1 < Standard 2 < Standard 3 < Standard 4 < Standard 5 < Standard 6.
3.2.- R-value of standard curve should be 0.900 - 0.999. R-value represent how well the experimental data fit the regression line. A significant difference in observed Ct values between replicates will lower the R-value.
3.3.- The standard curve slope result should be all negative.
3.4.- The desired amplification efficiencies vary from 90% to 110%. The theoretical maximum of 100% indicates that the polymerase enzyme is functioning at its maximum capacity. Low reaction efficiencies may be caused by poor primer design or by suboptimal reaction conditions. Reaction efficiencies >110 may indicate pipetting error in your serial dilutions or coamplification of nonspecific products, such as primer-dimers.

Visual explanation FAQ:

1.-Positive control:

The Positive control assay uses a FAM dye and should be detected through the Green channel of your real time PCR instrument **(see table 8).**
For copy number determination and as a positive control for the PCR set up, the kit contains a positive control template. This can be used to generate a standard curve of Spawner-isolated mortality virus copy number / Ct value.
Alternatively, the positive control can be used at a single dilution Spawner-isolated mortality virus on where full quantitative analysis of the sample is not required. Each time the kit is used, at least one positive control reaction must be included in the run. **Particularly, due to amount of this reagent, you should run a positive control for each 12 samples.** A positive result indicates that the primer and probes for detecting the target Spawner-isolated mortality virus gene worked properly in that particular experimental scenario. If a negative result is obtained the test results should be invalid and must be repeated **(see Table 10).** Sealing all other samples and negative controls before pipetting the positive control into the positive control well tube.

2.-Internal Control:

The internal control is included in Primer, Probes and Internal Control Mix along to the target pathogen detection. In order to interpreted results, read the yellow channel. The internal control assay uses a HEX dye and should be detected through the Yellow channel of your real time PCR instrument and gives a Ct value of 28 (+/-5) depending on the level of sample dilution and concentration. A positive result through the Yellow channel therefore indicates that PCR conditions are suitable for detection of the target pathogen gene. If a negative result is obtained through the Yellow channel the results should be analyzed by combination of result, follow the **Table 10** data.

3.-Negative control:

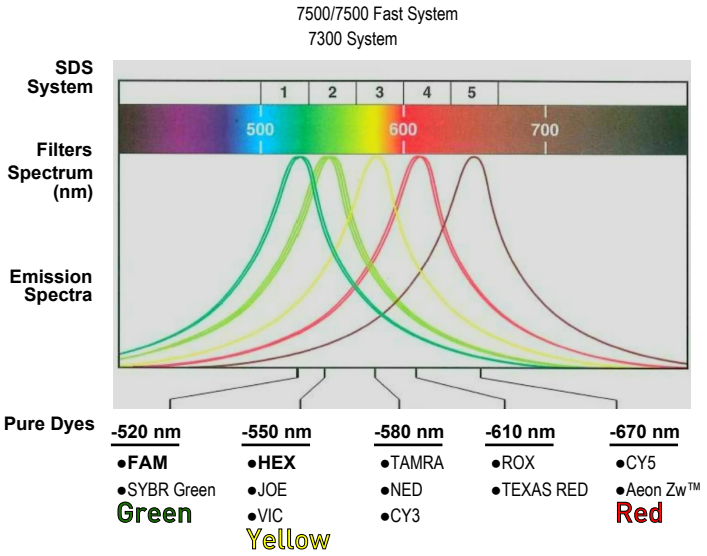
To confirm absence of contamination a negative control reaction should be included every time the kit is used. Particularly, due to amount of this reagent, you should run a negative control for each 12 samples. In this instance the PCR grade water should be used in place of template. A negative result indicates that the reagents have not become contaminated. If a positive result and Ct value less than 36 is obtained, the results should be analyzed and check if a correct amplification curve was obtained. When you obtain a clear amplification curve you should consider repeat your assay due to probably the sample was contaminated **(see Table 10)**.

* Remember: Run a positive control and negative control for each 12 samples. For reaction mix you must use Universal qPCR Master Mix.

*You must use quencher and reporter dye to setup your software **(see table 8)** and run the following channel:

Table 8. Fluorogenic probes, Channels and Dyes

Channel	Source	Detector	Dyes
Green	470 (Nm)	520 (Nm)	**FAM**, Sybr green1, Fluorescein, Eva green, Alerxa flour 488
Yellow	530 (Nm)	550 (Nm)	Joe, Vic, **HEX**, Tet, Cal Fluorgold 540, YaKima Yellow
Orange	585 (Nm)	610 (Nm)	Rox, Cal Fluor Red 610, Cy3.5, Texas Red, Alexa Fluor 568
Red	625 (Nm)	660 (Nm)	Cy5, Quasar 670, Lightcycler, Red 640, Alexa Fluor 633, Aeon Zw™
Crimson	680 (Nm)	710 (Nm)	Quasar 705, Lightcycler Red 705, Alexa Fluor 680



Important Note:

Probes for sample and controls mentioned in manuals are just a reference and it not imply that these probes will be the final fluorophores for a purchased kit. The probes combination will be depending of several factor and this information will be available in the Certificate of Analysis when you purchase one of our PCR Kits. FAM and HEX fluorophores are just referential.

Is important mentioned that we can develop special request for other pathogens or multiplex detection according client's specifications.

We strongly recommend don't use or combine our products with reagents from another kits or unknown provenance. We can't assure good result if incompatibility problems occur.

Temperature

All our reagents are made through protein engineering and are stable at room temperature, the label temperature is just a recommendation after the product is open.

Table 9. Store Temperature Kits

	Store temperature	The label temperature
	Shipping temperature	At room temperature

Table 10. Interpretation of Results

Sample	I.C	N.C	P.C	Result
+	+	-	+	POSITIVE
+	-	-	+	POSITIVE
+	+	+	+	Check Ct and Consider repeat assay*
+	-	+	+	Check Ct and Consider repeat assay*
+	-	-	-	NEGATIVE
+	+	-	-	NEGATIVE
-	+ / -	+ / -	+ / -	NEGATIVE

* * Sometimes amplification curves for Negative or Internal control with Ct < 30 is generate, but it's not necessary a Positive result. You should see and determinate if is a sigmoid curve. If the amplification curve isn't sigmoid you should consider as negative result.

Table 11. Products Specifications

Technology	5' nuclease probe based real time PCR assay
Type of nucleic acid Kit	DNA
Kit storage	Shipped at room temperature, the label temperature is just a recommendation after the product is open.
Detection Limit	See Quality Control file. **Request it!**
Sensitivity & Specify	Ct value between 12 – 36 should be taken positive. Value between 36-40 Ct should be taken as marginal positive. Ct above 40 must be considered as negative.
Controls included	Internal control, Positive control and Negative control included.
Channels	FAM Green channel detect pathogen amplicons. HEX Yellow channel detect internal control amplicons. FAM Green channel detect Positive Control.

Gentaur Molecular Products BVBA
Address: Voortstraat 49, 1910 Kampenhout, Belgium
T: 0032 16 58 90 45 | **E:** info@gentaur.com
Websites: www.gentaur.com | www.maxanim.com